



RESEARCH

Open Access

Enhanced photo-fermentative H₂ production using *Rhodobacter sphaeroides* by ethanol addition and analysis of soluble microbial products

Dong-Hoon Kim¹, Ji-Hye Lee¹, Seoktae Kang², Patrick C Hallenbeck³, Eui-Jin Kim⁴, Jeong K Lee⁴ and Mi-Sun Kim^{1,5*}

Abstract

Background: Biological fermentation routes can provide an environmentally friendly way of producing H₂ since they use renewable biomass as feedstock and proceed under ambient temperature and pressure. In particular, photo-fermentation has superior properties in terms of achieving high H₂ yield through complete degradation of substrates. However, long-term H₂ production data with stable performance is limited, and this data is essential for practical applications. In the present work, continuous photo-fermentative H₂ production from lactate was attempted using the purple non-sulfur bacterium, *Rhodobacter sphaeroides* KD131. As a gradual drop in H₂ production was observed, we attempted to add ethanol (0.2% v/v) to the medium.

Results: As continuous operation went on, H₂ production was not sustained and showed a negligible H₂ yield (< 0.5 mol H₂/mol lactate_{added}) within two weeks. Electron balance analysis showed that the reason for the gradual drop in H₂ production was ascribed to the increase in production of soluble microbial products (SMP_s). To see the possible effect of ethanol addition, a batch test was first conducted. The presence of ethanol significantly increased the H₂ yield from 1.15 to 2.20 mol H₂/mol lactate_{added}, by suppressing the production of SMPs. The analysis of SMPs by size exclusion chromatography showed that, in the later period of fermentation, more than half of the low molecular weight SMPs (< 1 kDa) were consumed and used for H₂ production when ethanol had been added, while the concentration of SMPs continuously increased in the absence of ethanol. It was found that the addition of ethanol facilitated the utilization of reducing power, resulting in an increase in the cellular levels of NAD⁺ and NADP⁺. In continuous operation, ethanol addition was effective, such that stable H₂ production was attained with an H₂ yield of 2.5 mol H₂/mol lactate_{added}. Less than 15% of substrate electrons were used for SMP production, whereas 35% were used in the control.

Conclusions: We have found that SMPs are the key factor in photo-fermentative H₂ production, and their production can be suppressed by ethanol addition. However, since external addition of ethanol to the medium represents an extra economic burden, ethanol should be prepared in a cost-effective way.

Keywords: Photo-fermentative hydrogen production, Electron balance, Soluble microbial products, Ethanol, Lactate, Size exclusion chromatography

* Correspondence: bmmskim@kier.re.kr

¹Biomass and Waste Energy Laboratory, Korea Institute of Energy Research, 152 Gajeong-ro, Daejeon, Yuseong-gu 305-343, Republic of Korea

⁵Division of Renewable Energy Engineering, University of Science and Technology, 217 Gajeong-ro, Daejeon, Yuseong-gu 305-350, Republic of Korea

Full list of author information is available at the end of the article

Background

Hydrogen (H_2), considered the cleanest energy alternative to fossil fuels, is now made exclusively by using fossil fuels: by steam reforming of natural gas, thermal cracking of light oil, and coal gasification. In contrast, biological fermentation routes can provide an environmentally friendly way of producing H_2 since they use renewable biomass as feedstock and proceed under ambient temperature and pressure [1,2]. Biological H_2 production occurs via two main pathways, classified as dark and photo-fermentation according to light dependency. Dark fermentative production has the advantage that the organic substrates are converted to H_2 at a fast production rate, with, however, the disadvantage that volatile fatty acids are produced as side products, thus limiting total H_2 yields [3,4].

Photo-fermentation is a process that can theoretically achieve maximum H_2 yields through complete degradation of substrates [5]. Purple non-sulfur (PNS) bacteria have been the most intensively studied photosynthetic H_2 -producing bacteria due to their demonstrated high substrate conversion yields. Under illuminated anaerobic conditions, PNS bacteria can grow photoheterotrophically using organic substrates and can potentially produce H_2 . However, they have several alternative metabolic modes such as photoautotrophy, fermentation, and aerobic/anaerobic respiration, which are non- H_2 -producing or even H_2 -consuming pathways [6]. Therefore, it is critical to adjust the culture conditions so that they are suitable for H_2 production.

Although PNS bacteria can use a wide variety of organic acids as a carbon source, the catabolic pathway involved depends on the substrate type. For example, acetate and butyrate are easily converted to acetyl units during metabolism by PNS bacteria and yield mainly polyhydroxyalkanoic acids (PHAs) rather than H_2 [7]. This leads to a decrease in H_2 production, since PHA production is an undesirable electron sink in competition with photo-fermentative H_2 production. Compared to acetate and butyrate, lactate and succinate are known to yield more H_2 [6,8]. Furthermore, since lactate can be easily obtained from anaerobic fermentation of agricultural and food wastes, it could be an ideal substrate for photo-fermentative H_2 production [9].

Along with PHAs, soluble microbial products (SMPs) can also be an electron sink in photo-fermentation, competing with the H_2 production pathway [10]. SMPs are soluble organic compounds that are released during biomass growth and cellular decay through normal metabolic processes [11,12]. Most studies on SMPs have been focused on aerobic systems, because SMPs often form the majority of the effluent chemical oxygen demand (COD) from biological treatment systems and are related to bio-fouling in membrane bioreactors (MBRs) [13,14]. However, there is little information about SMP production

during photo-fermentation. Yilmaz et al. [10] quantitatively analyzed the partitioning of nutrient electrons into H_2 , cell biomass, PHAs, and SMPs in photo-fermentation by *Rhodobacter sphaeroides*. They reported that SMPs were a major electron sink, taking a large amount of reducing power away from H_2 production. Similarly, Kim and Kim [15] found that 3% to 17% of substrate electrons were partitioned to SMP production during semi-continuous photo-fermentation, suggesting that SMPs are a major electron sink, taking large amounts of reducing power away from H_2 production.

So far, numerous studies have been conducted to optimize photo-fermentative H_2 production in batch-type operations, but there are only a few reports on continuous operation [16-18]. To our knowledge, long-term H_2 production data with stable performance is limited; however, this data would be essential for the practical application of the process. In the present work, continuous photo-fermentative H_2 production from lactate was attempted using the PNS bacterium *Rhodobacter sphaeroides* KD131. Electron balances were established to evaluate electron partitioning of lactate into H_2 , cell biomass, and SMPs. As a gradual drop in H_2 production was observed during continuous operation, 0.2% (v/v) ethanol was externally added to the medium, since it has recently been shown that the presence of ethanol can enhance H_2 production by *R. sphaeroides* [19]. In addition, size exclusion chromatography (SEC) and excitation-emission matrix (EEM) analysis were carried out to characterize the SMPs produced during photo-fermentation.

Results and discussion

Continuous performance

As shown in Figure 1a, H_2 production from lactate was not sustained. H_2 production gradually dropped and showed a negligible H_2 yield (< 0.5 mol H_2 /mol lactate_{added}) within two weeks. The theoretical maximum H_2 yield from lactate is 6 mol H_2 /mol lactate_{added} [7]. Although the fermenter was reseeded with a new inoculum (trials 2 and 3), the same phenomenon was observed. Unlike H_2 production, lactate degradation and cell growth did not fluctuate during the entire operation period, suggesting that some portion of electrons contained in the substrate was diverted towards other paths (Figure 1b). Here, we have quantified the SMPs and demonstrate that changes in SMP concentration show an opposite trend to H_2 production.

Establishing an electron balance is important in anaerobic fermentative processes, as it indicates the accuracy of the experiment and provides basic information regarding metabolic flux control [15,20,21]. In the present study, the COD was used as the common unit for electron balance, and the substrate and all metabolic products were converted to this unit. The electrons in the

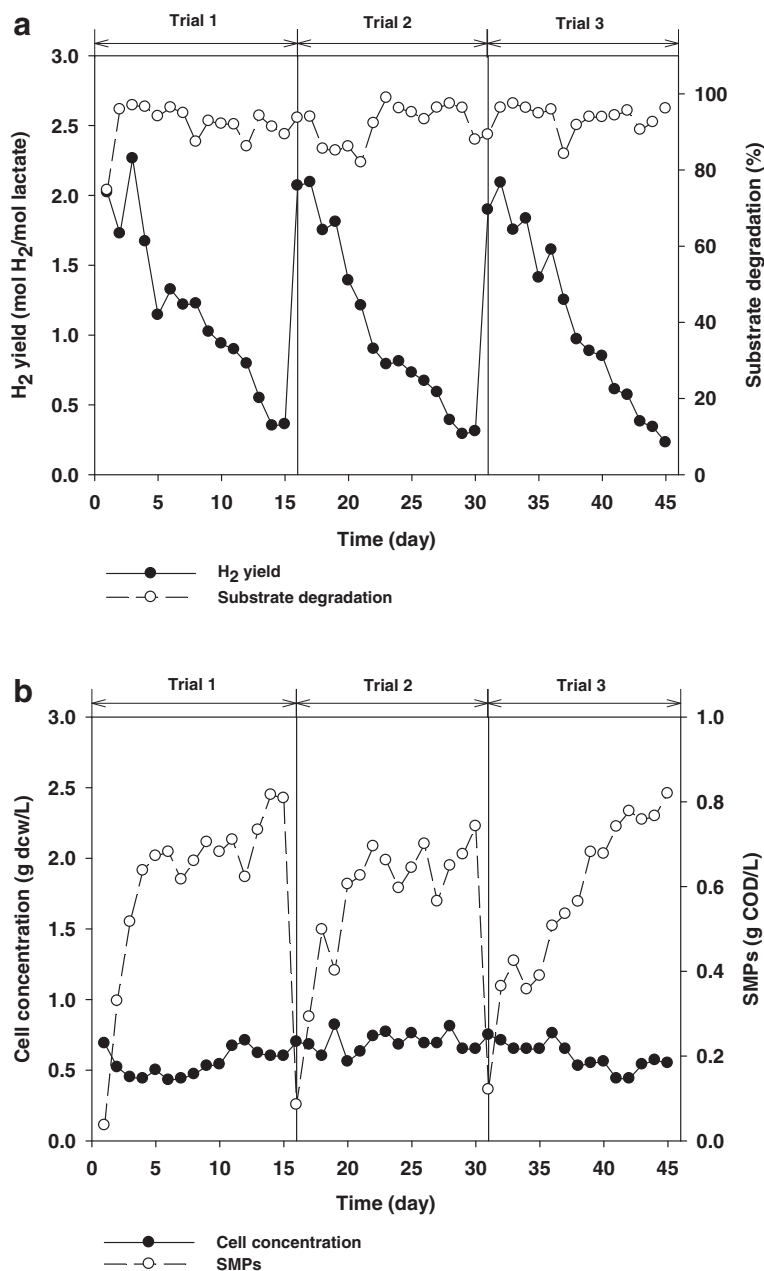


Figure 1 Daily performance of photo-fermentative H_2 production from lactate. (a) H_2 yield and lactate degradation, and (b) cell concentration and SMPs.

substrate that were utilized can be distributed to H_2 , cell growth, and SMPs. The result of this electron balance exercise can be seen in Figure 2. Unlike the work of Yilmaz et al. [10], in our study, PHA was not separately considered with cell growth since most of the PHA is retained inside the cell [22]. The amount of SMPs was measured by subtracting the residual lactate from the COD of samples that had been filtered through a 0.45- μ m membrane. The COD of a cell was calculated by assuming a composition of $C_5H_7O_2N$, resulting in a COD

value of 1.42 g COD/g DCW (dry cell weight) [23]. While a fully accurate and precise electron balance was not possible since H_2 production and SMP concentration fluctuated during the experimental period, it was informative to roughly determine electron distribution at the point of failure by collecting data at each trial after 10 days of operation.

The sum of metabolic products and the residual substrate accounted for $100 \pm 10\%$ of the input substrate, which indicates that the experiments were conducted

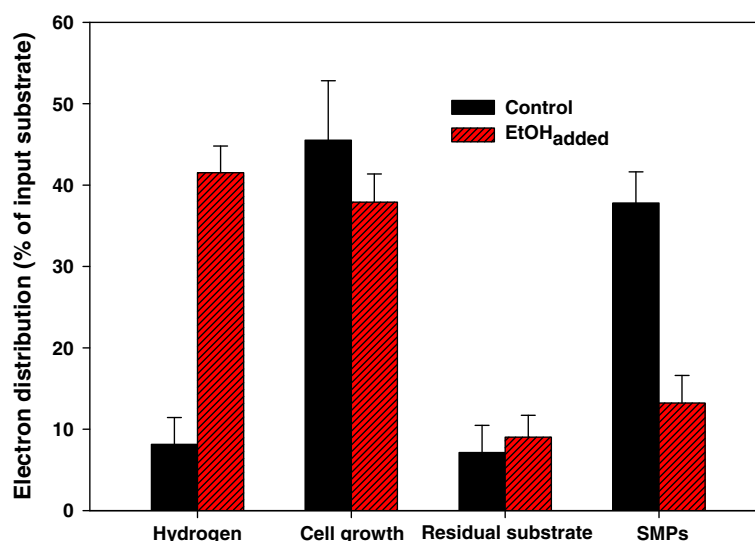


Figure 2 Effect of ethanol addition on electron distribution in continuous photo-fermentative H₂ production. In the control (absence of ethanol), data from three independent experiments were collected after 10 days of operation and were averaged.

with sufficient accuracy. From the electron balance obtained, a number of observations about the reasons for unsuccessful H₂ production can be made. In the control, it would seem that most of the electrons were consumed for cell growth and SMP production rather than for H₂ production. Around 45% and 38% of the electrons were diverted toward cell growth and SMP production, respectively. As the cell concentration did not vary during the continuous operation, the reason for the gradual drop in H₂ production can be ascribed to the increase in SMP production as continuous operation proceeded.

Effect of ethanol addition on batch-type fermentation

We first assessed any possible effect of ethanol addition (0.2% v/v) by examining batch-type photo-fermentative H₂ production (Figure 3). In the presence of ethanol, H₂ production continued for 80 h while there was no more H₂ production after 40 h in the control. Moreover, H₂ yield was enhanced from 1.15 to 2.20 mol H₂/mol lactate_{added} by the addition of ethanol (448 mL H₂/L-broth = 1 mol H₂/mol lactate_{added}). There was not much difference in the profile of residual lactate concentration and pH (data not shown), but significantly different patterns were observed for cell concentration and SMP production depending on whether ethanol had been added or not. As operation continued, the pH gradually increased from 7.5 to 8.0 with lactate consumption. In the control (in the absence of ethanol), the cell concentration increased significantly until 20 h, and then gradually decreased afterwards. However, in the presence of ethanol, cell concentration continuously increased, finally reaching about a 20% higher cell concentration. In contrast, while SMP production continued for the entire

experimental period in the control, it can be clearly seen that the addition of ethanol suppressed SMP production after 40 h. It is likely that the production of SMPs in the later periods of incubation resulted from microbial decay, since no further lactate degradation occurred after 50 h.

Characterization of SMPs

SMPs are defined as soluble organic compounds that are released during normal substrate metabolism (utilization-associated SMPs, UAPs) and decay in biological processes (biomass-associated SMPs, BAPs) [11]. UAPs are known to be further utilized by microorganisms, while BAPs are responsible for final effluent organic matter [11,24]. During batch photo-fermentation, it is likely that cell decay (or production of BAPs) became dominant after 40 h in the absence of ethanol, while additional conversion of SMPs to H₂ took place in the presence of ethanol, as indicated by the data shown in Figure 4. Indeed, characterization of SMPs strongly supports this idea. The molecular weight distribution of SMPs analyzed at various incubation times showed that the mass of high molecular weight SMPs kept increasing during the incubation in both samples. However, the mass of low molecular weight SMPs (< 1 kDa) was significantly reduced from 197 mg dissolved organic carbon (DOC)/L at 32 h fermentation to 96 mg DOC/L at 72 h fermentation in the presence of 0.2% ethanol, while that of the control increased from 205 mg DOC/L to 280 mg DOC/L over the same time period. This supports the contention that the low molecular weight SMPs were converted to H₂ in the presence of ethanol.

In Figure 5, the EEM of SMPs taken at various fermentation times shows that the locations of peaks in the two samples are similar except for the peak with excitation at

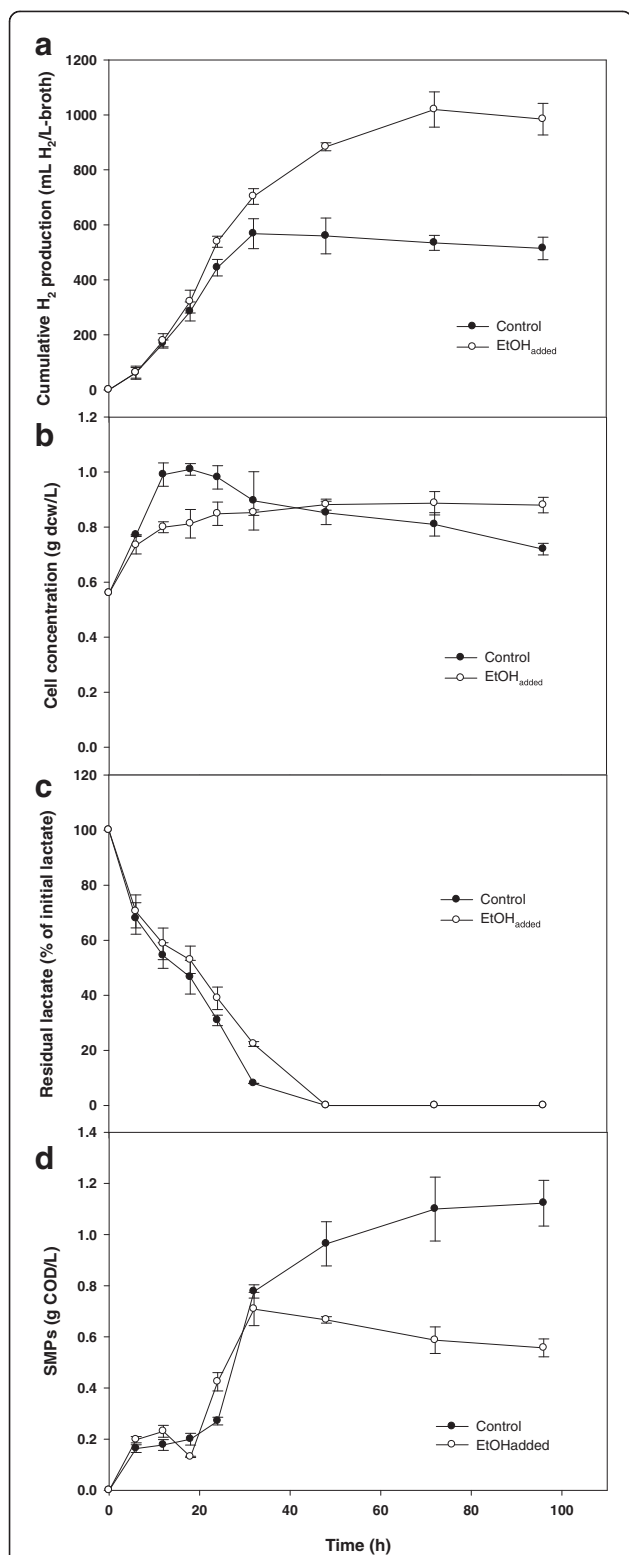


Figure 3 Effect of ethanol addition on the batch performance of photo-fermentative H₂ production from lactate. (a) H₂ production, (b) cell concentration, (c) residual lactate, and (d) SMPs.

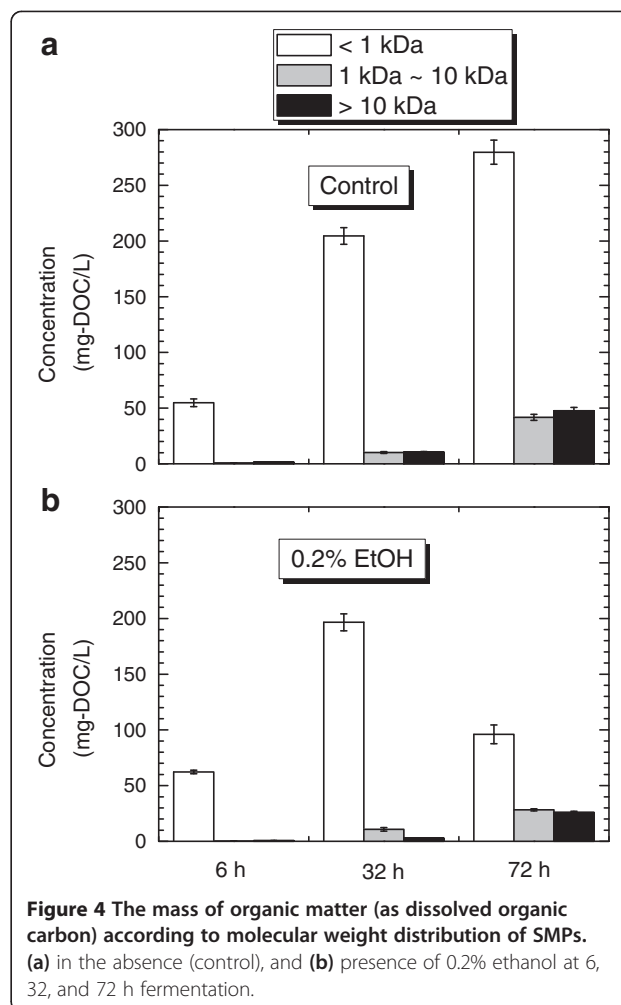


Figure 4 The mass of organic matter (as dissolved organic carbon) according to molecular weight distribution of SMPs. (a) in the absence (control), and (b) presence of 0.2% ethanol at 6, 32, and 72 h fermentation.

around 270 nm and emission at around 340 nm, thought to correspond to BAPs [25]. In the presence of ethanol, the BAP peaks were weaker than those of the control, indicating that the majority of SMPs produced in the presence of ethanol were mostly UAPs which could be further fermented to produce H₂. After 72 h of incubation, at which point photo-fermentation is complete in both samples, the peaks responsible for high molecular weight SMPs (> 10 kDa) were revealed to be due to humic acid-like (excitation-emission at 320 to 360 nm and 400 to 450 nm) compounds which cannot be degraded by the micro-organism. Compared to the control, these peaks were also weaker in the EtOH_{added} case, suggesting that the presence of ethanol can lead to less production of SMPs.

Alteration of redox balance by the addition of ethanol

The reducing power for nitrogenase activity is provided by ferredoxin, which is reduced by NADPH/NADP⁺ (nicotinamide adenine dinucleotide phosphate):ferredoxin oxidoreductase [26]. Thus, the H₂ production by nitrogenase is affected by the state of cellular reducing equivalents. To

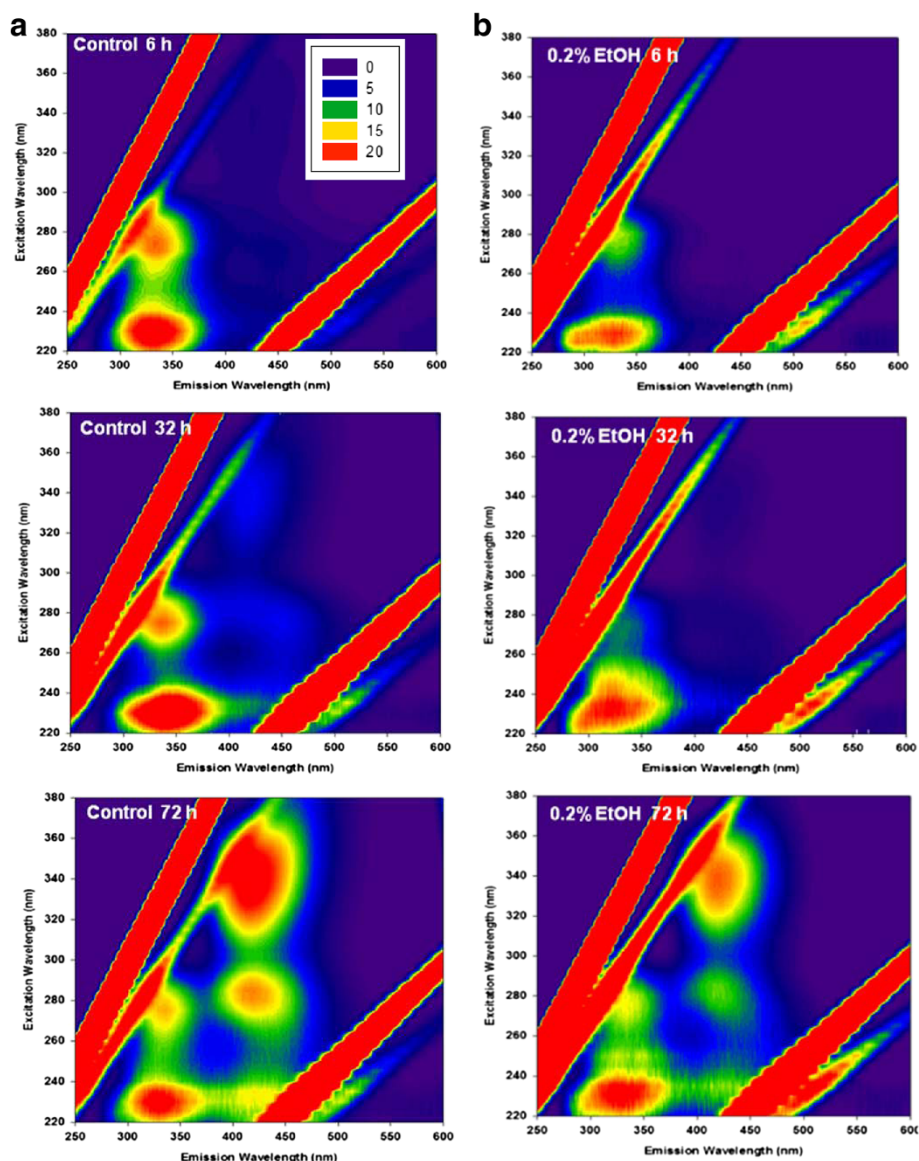


Figure 5 Excitation-emission matrix of soluble microbial products. (a) in the absence (control), and (b) presence of 0.2% ethanol after 6, 32, and 72 h fermentation.

investigate the effect of ethanol on the equilibrium of cellular reducing equivalents, the cellular levels of NAD(H) (nicotinamide adenine dinucleotide) and NADP(H) were determined in the absence and presence of 0.2% ethanol. As shown in Figure 6a, NAD⁺ was increased by up to approximately 50% with the addition of ethanol, whereas the level of NADH was relatively unchanged by the presence of ethanol, in comparison with the case of the control. Interestingly, concomitant with the 10% to 30% decrease in NADPH, NADP⁺ increased almost threefold by the addition of ethanol (Figure 6b). Accordingly, the ratios of both NADH/NAD⁺ and NADPH/NADP⁺ were lowered to 40% to 70% of the control values that were determined in the absence of ethanol (Figure 6c). Since the nitrogenase

activity of *R. sphaeroides* is elevated in the presence of ethanol [19], the consumption of reducing power appears to be accelerated by ethanol. Previously, an increase in the cellular NAD⁺ level was also observed following the deletion of nonessential metabolic pathways in *R. sphaeroides* [27]. When the metabolic flux leading to H₂ production is increased, the equilibrium of cellular reducing equivalents appears to move towards a more oxidized state.

Excessive reducing power is generated during photosynthetic growth on reduced carbon compounds, and balance needs to be maintained through reductive pathways such as CO₂ fixation, thereby yielding NAD⁺ and NADP⁺ [28]. It has been suggested that, when growing heterotrophically under photosynthetic conditions, *R. sphaeroides* forms

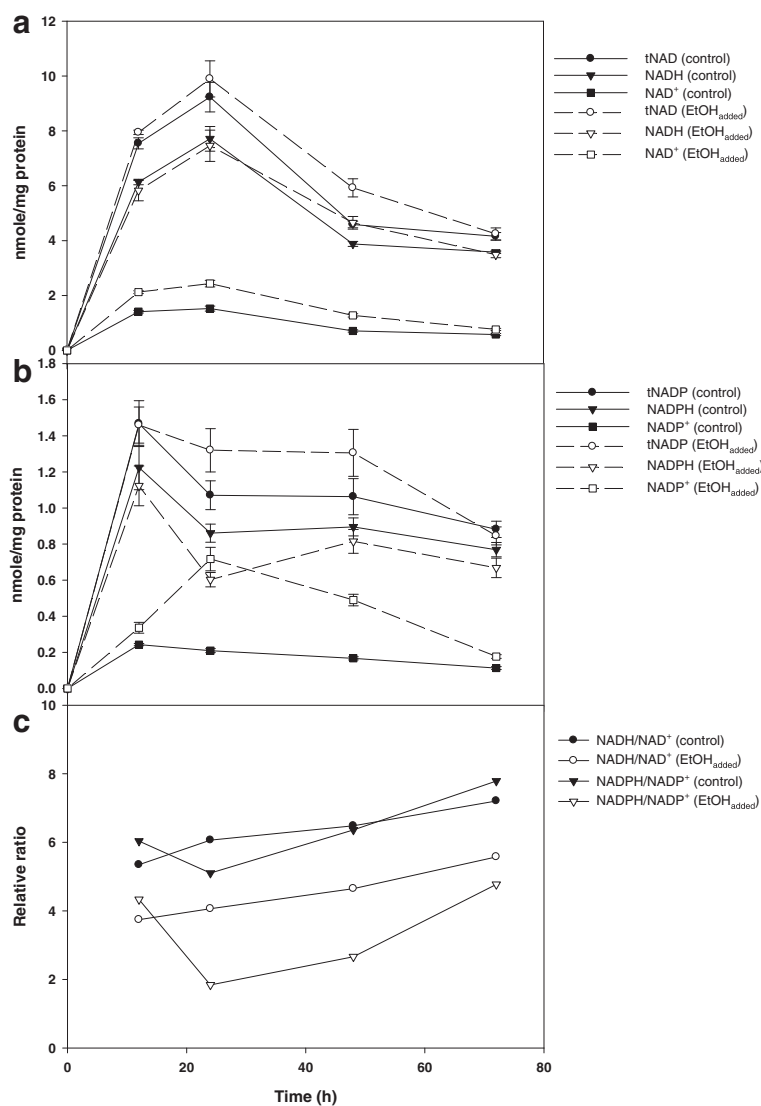


Figure 6 Cellular level of NAD(H) and NADP(H) in the absence and presence of 0.2% ethanol of batch-type photo-fermentation. (a) NADH, NAD⁺, and the total level of NADH + NAD⁺ (tNAD), **(b)** NADPH, NADP⁺, and the total level of NADPH + NADP⁺ (tNADP), and **(c)** the ratios of both NADH/NAD⁺ and NADPH/NADP⁺.

SMPs to remove excess reducing power [10]. As can be seen in Figure 6, it turns out that the NAD(H) and NADP(H) pools in the control photosynthetically grown *R. sphaeroides* still remain in the reduced state. Since BAPs were dominantly produced under photosynthetic conditions, the formation of BAPs is likely to result from this reduced state. On the other hand, when ethanol was added to the medium, the redox balance shifted to a more oxidized state, implying that more reducing power had been consumed by the cell. The level of BAPs was consistently decreased in the presence of ethanol. Moreover, UAPs, which were produced instead of BAPs in cultures exposed to ethanol, can be further utilized as a secondary fermentative organic source for H₂ production. Thus,

although excess reducing power can be eliminated by SMP formation (more specifically, BAP formation) during photo-fermentative growth of *R. sphaeroides*, more reducing power appears to be used up in the presence of ethanol, resulting in an increase of cellular NAD⁺ and NADP⁺.

Effect of ethanol addition on continuous operation

Next we assessed the effect of ethanol addition on continuous photo-fermentative H₂ production from lactate. Ethanol (0.2%) was added to the medium, which was continuously fed to the fermenter. As shown in Figure 7a, H₂ production was stable for one month while giving an average H₂ yield of 2.5 mol H₂/mol lactate_{added} (corresponding to 41.6% of the total electron consumption). Lactate

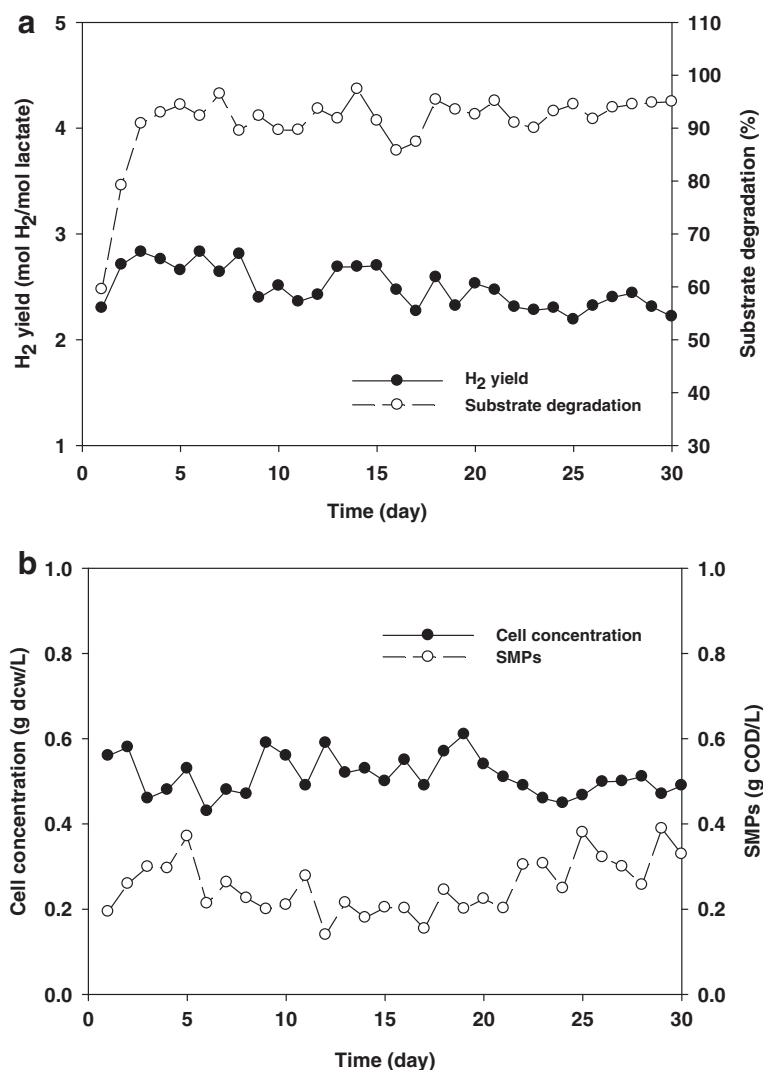


Figure 7 Performance of continuous photo-fermentative H₂ production from lactate in the presence of 0.2% ethanol. (a) H₂ production and substrate degradation and (b) cell concentration and SMPs.

degradation and cell concentration were stable over this time period, and SMP production was minimized (Figure 7b). Less than 15% of electrons contained in the substrate were used for SMP production, whereas 35% were used in the control (Figure 2).

From the series of experiments, we found that SMPs are the key factor in photo-fermentative H₂ production, and the SMP production can be suppressed by ethanol addition. However, since the external addition of ethanol to the medium represents an extra economic burden, ethanol should be prepared in a cost-effective way. For example, hetero-fermentative lactic acid bacteria ferment organic wastes to lactate and ethanol, which could be a suitable feedstock for photo-fermentative H₂ production [29]. This process could be further developed by the optimization of operational parameters such as cell

retention time, organic loading rate, and pH, all of which could influence SMP production.

Conclusions

From the series of batch and continuous experiments on photo-fermentative H₂ production from lactate in the absence and presence of ethanol presented here, the following conclusions can be drawn:

1. H₂ production was not sustained during continuous photo-fermentative H₂ production, as the H₂ yield dropped lower than 0.5 mol H₂/mol lactate_{added} within two weeks with increasing production of SMPs. Rather than H₂, most of the electrons were diverted towards cell growth and SMP production.
2. In batch operation, the presence of ethanol (0.2%) increased the H₂ yield from 1.15 to 2.20 mol H₂/mol lactate_{added} by suppressing the production of

SMPs. Cell decay became dominant after 40 h in the absence of ethanol, while additional conversion of SMPs to H₂ took place in the presence of ethanol. In the later period of fermentation, more than half of the low molecular weight SMPs (< 1 kDa) were consumed when ethanol had been added, whereas the SMP concentration continuously increased in the absence of ethanol. It was found that the addition of ethanol facilitated the utilization of reducing power, resulting in an increase in the cellular level of NAD⁺ and NADP⁺.

3. When ethanol was added during continuous operation, H₂ production was stable for one month with an average H₂ yield of 2.5 mol H₂/mol lactate_{added} (corresponding to 41.6% of total electron consumption). Less than 15% of substrate electrons were used for SMPs production, whereas 35% were used in the control.

Methods

Inoculum preparation

The phototrophic bacterium *R. sphaeroides* KD131, isolated from mud off the coast of Daebu Island in the West Sea of South Korea, was used for photo-fermentative H₂ production. *R. sphaeroides* KD131 was pre-cultured in a modified Siström's broth [30] containing 4 mM (NH₄)₂SO₄, 0.3 mM L-aspartic acid, and 20 mM lactate at 30°C for 24 h under 110 W/m² irradiance using halogen lamps (12 V, 50 W). The cells were capped in anaerobic tubes with O-rings and collected by centrifugation (8,000 rpm for 10 min, Supra 22 K, Hanil Co.) under anaerobic conditions, and used as an inoculum for H₂ production.

Experiments

For continuous operation, a 3.5-L glass fermenter (working volume of 3.0 L, 830 mm high by 80 mm in diameter) installed with a pH sensor at the top was used. Centrifuged biomass was added to reach an initial cell concentration of 0.56 g DCW/L equivalent to an optical density of 1.0. After purging with Ar gas (99.999%) for 1 h, the fermenter was operated for 48 h by batch mode as an adaptation period, and then switched to continuous mode. One liter of lactate (20 mM) containing medium (a modified Siström's broth containing 4 mM (NH₄)₂SO₄ and 0.3 mM L-aspartic acid) was continuously fed and removed per day, corresponding to three days of hydraulic retention time (HRT). During the operation, pH was maintained at 7.5 ± 0.2 by use of the pH sensor and the addition of 1 N HCl solution.

For the batch experiments to assess the effect of ethanol addition (0.2% v/v) on H₂ production, 100 mL (effective volume of 50 mL) serum bottles were used, and the preparation procedure was the same as that used for continuous operation. The initial substrate concentration and cell concentration were 20 mM lactate and 0.56 g DCW/L, respectively. The amount of H₂ production

and the concentration of cell, residual lactate, and SMPs were measured periodically at intervals of 5 to 20 h. The experiment was conducted in triplicate and the results were averaged.

The light intensity (measured at the surface of the fermenter) was adjusted to 110 W/m² using halogen lamps (12 V, 50 W), and all experiments were conducted in a temperature-controlled room at 30°C.

Analytical methods

Measured biogas production was corrected to standard temperature (0°C) and pressure (760 mmHg) (STP). The H₂ content in the biogas was analyzed using a gas chromatograph (Model 14-B, Shimadzu Co., Japan) equipped with a thermal conductivity detector and a stainless steel column packed with Molecular Sieve 5A (80/200 mesh; Altech, Deerfield, USA). 1 mL of sampled gas was injected, and the lower detection limit was 0.1%. The temperatures of the injector, detector, and column were kept at 100, 120, and 80°C, respectively. Organic acids were analyzed using a high performance liquid chromatograph (HPLC, Model VP, Shimadzu Co., Japan) equipped with a sulfonated divinyl benzene-styrene co-polymer column (300 mm × 7.8 mm, Aminex HPX-87H, BioRad, USA). 20 µL of sampled liquid was injected, and the lower detection limit was 0.01 mM. The column temperature was maintained at 35°C, and a photometric detector (216 nm) was used to quantify the organic acids eluted from the column. An aqueous solution of 10 mM H₂SO₄ was used as the eluting buffer and was dispensed at a flow rate of 0.6 mL/min. Liquid samples were pretreated with a 0.45-µm membrane filter (Millipore, USA) prior to injection to HPLC. The COD concentration was measured according to standard methods [31].

The supernatant filtered through a 0.45-µm membrane was regarded as SMPs after subtracting the residual lactate. After normalization of DOC to 3 mg DOC/L, the EEMs were examined using fluorescent spectroscopy (Shimadzu RF530, Japan) at excitation from 220 nm to 380 nm and emission from 250 nm to 600 nm. The EEMs were further normalized by using signals from water molecules as well as Rayleigh scattering, as previously suggested [25]. The average molecular weight of SMPs was analyzed by size exclusion chromatography using an HPLC (Younglin YL9101, Korea; Waters column, USA) with polysaccharides (dextran) as the molecular weight standard. The measurements of cellular NAD⁺, NADH, and their ratio as well as NADP⁺, NADPH, and their ratio were determined by using an NAD⁺/NADH quantification kit (BioVision, USA) and an NADP⁺/NADPH quantification kit (BioVision, USA), respectively.

Abbreviations

BAPs: biomass-associated soluble microbial products; COD: chemical oxygen demand; DCW: dry cell weight; DOC: dissolved organic carbon;

EEM: excitation-emission matrix; HPLC: high performance liquid chromatograph; HRT: hydraulic retention time; MBRs: membrane bioreactors; NAD: nicotinamide adenine dinucleotide; NADP: nicotinamide adenine dinucleotide phosphate; PHA: polyhydroxyalkanoic acid; PNS: purple non-sulfur; SEC: size exclusion chromatography; SMPs: soluble microbial products; STP: standard temperature and pressure; UAPs: utilization-associated soluble microbial products.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

DHK performed the experiments, analyzed the results, and drafted the manuscript. JHL, SK, and EJK participated in the experiments. PCH and JKL commented on the manuscript. M-SK coordinated the study and revised the manuscript. All authors approved the final manuscript.

Acknowledgements

This work was supported by the Intelligent Synthetic Biology Center of Global Frontier Project funded by the Ministry of Education, Science and Technology (2011-0031955), and was conducted under the framework of the Research and Development Program of the Korea Institute of Energy Research (KIER) (B4-2434-04).

Author details

¹Biomass and Waste Energy Laboratory, Korea Institute of Energy Research, 152 Gajeong-ro, Daejeon, Yuseong-gu 305-343, Republic of Korea. ²Department of Civil Engineering, Kyung Hee University, 1732 Deokyoungdaero, Yongin, Giheung, Gyeonggi-do 446-701, Republic of Korea. ³Departement of Microbiology and Immunology, University of Montreal, CP 6128, Succursale Centre-Ville, Montreal, QC H3C 3J7, Canada. ⁴Department of Life Science and Basic Science Institute for Cell Damage Control, Sogang University, Mapo, Shin-su 1, Seoul 121-742, Republic of Korea. ⁵Division of Renewable Energy Engineering, University of Science and Technology, 217 Gajeong-ro, Daejeon, Yuseong-gu 305-350, Republic of Korea.

Received: 9 January 2014 Accepted: 9 May 2014

Published: 27 May 2014

References

- Hawkes FR, Hussy I, Kyazze G, Dinsdale R, Hawkes DL: Continuous dark fermentative hydrogen production by mesophilic microflora: principles and progress. *Int J Hydrogen Energy* 2007, **32**:172-184.
- Lee HS, Vermaas WFJ, Rittmann BE: Biological hydrogen production: prospects and challenges. *Trends Biotechnol* 2010, **28**(5):262-271.
- Kim DH, Kim MS: Hydrogenases for biological hydrogen production. *Int J Hydrogen Energy* 2011, **102**:8423-8431.
- Levin DB, Chahine R: Challenges for renewable hydrogen production from biomass. *Int J Hydrogen Energy* 2010, **35**:4962-4969.
- Chen CY, Liu CH, Lo YC, Chang JS: Perspectives on cultivation strategies and photobioreactor designs for photo-fermentative hydrogen production. *Bioresour Technol* 2011, **102**:8484-8492.
- Koku H, Eroglu I, Gunduz U, Yucel M, Turker L: Aspects of the metabolism of hydrogen production by *Rhodobacter sphaeroides*. *Int J Hydrogen Energy* 2002, **27**:1315-1329.
- Kim DH, Son H, Kim MS: Effect of substrate concentration on continuous photo-fermentative hydrogen production from lactate using *Rhodobacter sphaeroides*. *Int J Hydrogen Energy* 2012, **37**:15483-15488.
- Kim MS, Kim DH, Cha J: Culture conditions affecting H₂ production by phototrophic bacterium *Rhodobacter sphaeroides* KD131. *Int J Hydrogen Energy* 2012, **37**:14055-14061.
- Anam K, Habibi MS, Harwati TU, Susilaningih D: Photofermentative hydrogen production using *Rhodobium marinum* from bagasse and soy sauce wastewater. *Int J Hydrogen Energy* 2012, **37**(20):15436-15442.
- Yilmaz LS, Kontur WS, Sanders AP, Sohmen U, Donohue TJ, Noguera DR: Electron partitioning during light- and nutrient-powered hydrogen production by *Rhodobacter sphaeroides*. *Bioenerg Res* 2010, **3**:55-66.
- Ni BJ, Rittmann BE, Yu HQ: Soluble microbial products and their implications in mixed culture biotechnology. *Trends Biotechnol* 2011, **29**:454-463.
- Ni BJ, Zeng RJ, Fang F, Xie WM, Sheng GP, Yu HQ: Fractionating soluble microbial products in the activated sludge process. *Wat Res* 2010, **44**:2292-2302.
- Zuthi MFR, Ngo HH, Guo WS, Zhang J, Liang S: A review towards finding a simplified approach for modelling the kinetics of the soluble microbial products (SMP) in an integrated mathematical model of membrane bioreactor (MBR). *Int Biodeter Biodegr* 2013, **85**:466-473.
- Menniti A, Morgenroth E: Mechanisms of SMP production in membrane bioreactors: choosing an appropriate mathematical model structure. *Wat Res* 2010, **44**:5240-5251.
- Kim DH, Kim MS: Semi-continuous photo-fermentative H₂ production by *Rhodobacter sphaeroides*: effect of decanting volume ratio. *Bioresour Technol* 2012, **103**:481-483.
- Ren NQ, Liu BF, Zheng GX, Xing DF, Zhao X, Guo WQ, Ding J: Strategy for enhancing photo-hydrogen production yield by repeated fed-batch cultures. *Int J Hydrogen Energy* 2009, **34**:7579-7584.
- Lo YC, Chen CY, Lee CM, Chang JS: Photo fermentative hydrogen production using dominant components (acetate, lactate and butyrate) in dark fermentation effluents. *Int J Hydrogen Energy* 2011, **36**(21):14059-14068.
- Ozmihci S, Kargi F: Bio-hydrogen production by photo-fermentation of dark fermentation effluent with intermittent feeding and effluent removal. *Int J Hydrogen Energy* 2010, **35**:6674-6680.
- Oh EK, Kim EJ, Hwang HJ, Tong X, Nam JM, Kim MS, Lee JK: The photoheterotrophic H₂ evolution of *Rhodobacter sphaeroides* is enhanced in the presence of ethanol. *Int J Hydrogen Energy* 2012, **37**:15886-15892.
- Kim DH, Han SK, Kim SH, Shin HS: Effect of gas sparging on continuous fermentative hydrogen production. *Int J Hydrogen Energy* 2006, **31**:2158-2169.
- Lalman JA, Chaganti SR, Moon C, Kim DH: Elucidating acetogenic H₂ consumption in dark fermentation using flux balance analysis. *Bioresour Technol* 2013, **146**:775-778.
- Cetin D, Gunduz U, Eroglu I, Yucel M, Turker L: Poly-β-hydroxybutyrate accumulation and releasing by hydrogen producing bacteria, *Rhodobacter sphaeroides* O.U.001. A transmission electron microscopic study. *Afr J Biotechnol* 2006, **5**(22):2069-2072.
- Fang HHP, Liu H: Effect of pH on hydrogen production from glucose by a mixed culture. *Bioresour Technol* 2002, **82**:87-93.
- Abdelmelek SB, Greaves J, Ishida KP, Cooper WJ, Song W: Removal of pharmaceutical and personal care products from reverse osmosis retentate using advanced oxidation processes. *Environ Sci Technol* 2011, **45**(8):3665-3671.
- Chen W, Westerhoff P, Leenheer JA, Booksh K: Fluorescence excitation-emission matrix regional integration to quantify spectra for dissolved organic matter. *Environ Sci Technol* 2003, **37**:5701-5710.
- Jouanneau Y, Meyer C, Naud I, Klipp W: Characterization of an *fdxN* mutant of *Rhodobacter capsulatus* indicates that ferredoxin I serves as electron donor to nitrogenase. *Biochim Biophys Acta* 1995, **1232**:33-42.
- Tao Y, Liu D, Yan X, Zhou Z, Lee JK, Yang C: Network identification and flux quantification of glucose metabolism in *Rhodobacter sphaeroides* under photoheterotrophic H₂-producing conditions. *J Bacteriol* 2012, **194**:274-283.
- Laguna R, Tabita FR, Alber BE: Acetate-dependent photoheterotrophic growth and the differential requirement for the Calvin-Benson-Bassham reductive pentose phosphate cycle in *Rhodobacter sphaeroides* and *Rhodospseudomonas palustris*. *Arch Microbiol* 2011, **193**:151-154.
- Poolman B: Energy transduction in lactic acid bacteria. *FEMS Microbiol Rev* 1993, **12**:125-148.
- Sistrom WR: A requirement for sodium in the growth of *Rhodospseudomonas sphaeroides*. *J Gen Microbiol* 1960, **22**:778-785.
- American Public Health Association (APHA), American Water Works Association (AWWA), Water Environment Federation (WEF): *Standard methods for the examination of water and wastewater*. 20th edition. Washington, DC: APHA, AWWA, WEF; 1998.

doi:10.1186/1754-6834-7-79

Cite this article as: Kim et al.: Enhanced photo-fermentative H₂ production using *Rhodobacter sphaeroides* by ethanol addition and analysis of soluble microbial products. *Biotechnology for Biofuels* 2014 **7**:79.